



Providing Choice & Value

Generic CT and MRI Contrast Agents



CONTACT REP

AJNR

This information is current as of July 30, 2025.

Iron Deposition on SWI-Filtered Phase in the Subcortical Deep Gray Matter of Patients with Clinically Isolated Syndrome May Precede Structure-Specific Atrophy

J. Hagemeier, B. Weinstock-Guttman, N. Bergsland, M. Heininen-Brown, E. Carl, C. Kennedy, C. Magnano, D. Hojnacki, M.G. Dwyer and R. Zivadinov

AJNR Am J Neuroradiol 2012, 33 (8) 1596-1601

doi: <https://doi.org/10.3174/ajnr.A3030>

<http://www.ajnr.org/content/33/8/1596>

ORIGINAL
RESEARCH

J. Hagemeyer
B. Weinstock-Guttman
N. Bergsland
M. Heininen-Brown
E. Carl
C. Kennedy
C. Magnano
D. Hojnacki
M.G. Dwyer
R. Zivadinov



Iron Deposition on SWI-Filtered Phase in the Subcortical Deep Gray Matter of Patients with Clinically Isolated Syndrome May Precede Structure-Specific Atrophy

BACKGROUND AND PURPOSE: Increasing evidence suggests that iron deposition is present in the later stages of MS. In this study we examined abnormal phase values, indicative of increased iron content on SWI-filtered phase images of the SDGM in CIS patients and HC. We also examined the association of abnormal phase with conventional MR imaging outcomes at first clinical onset.

MATERIALS AND METHODS: Forty-two patients with CIS (31 female, 11 male) and 65 age and sex-matched HC (41 female, 24 male) were scanned on a 3T scanner. Mean age was 40.1 (SD = 10.4) years in patients with CIS, and 42.8 (SD = 14) years in HC, while mean disease duration was 1.2 years (SD = 1.3) in patients with CIS. MP-APT, NPTV, and normalized volume measurements were derived for all SDGM structures. Parametric and nonparametric group-wise comparisons were performed, and associations were determined with other MR imaging metrics.

RESULTS: Patients with CIS had significantly increased MP-APT ($P = .029$) and MP-APT volume ($P = .045$) in the pulvinar nucleus of the thalamus compared with HC. Furthermore, the putamen ($P = .004$), caudate ($P = .035$), and total SDGM ($P = .048$) displayed significant increases in MP-APT volume, while MP-APT was also significantly increased in the putamen ($P = .029$). No global or regional volumetric MR imaging differences were found between the study groups. Significant correlations were observed between increased MP-APT volumes of total SDGM, caudate, thalamus, hippocampus, and substantia nigra with white matter atrophy and increased T2 lesion volume ($P < .05$).

CONCLUSION: Patients with CIS showed significantly increased content and volume of iron, as determined by abnormal SWI-phase measurement, in the various SDGM structures, suggesting that iron deposition may precede structure-specific atrophy.

ABBREVIATIONS: CIS = clinically isolated syndrome; EDSS = Expanded Disability Status Scale; ETL = echo-train length; FIRST = fMRI-integrated registration and segmentation tool; Gd = gadolinium; GM = gray matter; HC = healthy controls; LV = lesion volume; MP-APT = mean phase of the abnormal phase tissue; NBV = normalized brain volume; NGMV = normalized gray matter volume; NLVV = normalized lateral ventricle volume; NPTV = normal phase tissue volume; NWMV = normalized white matter volume; pFOV = phase FOV; RRMS = relapsing-remitting MS; SDGM = subcortical deep GM

It has become increasingly clear that GM damage is present in patients with MS and involves both the cortical GM and SDGM structures.¹ GM damage is most extensive in patients with progressive MS,² but is already present in those with CIS³⁻¹⁰ and can predict conversion to clinically definite MS.¹¹⁻¹³

Most authors investigating iron deposition in MS have used imaging techniques such as T2 hypointensity,¹⁴⁻¹⁶ relaxometry,¹⁷⁻¹⁹ magnetic field correlation,²⁰ and SWI.²¹⁻²³ Accumulation of iron in the SDGM has also been observed histologically in MS,^{24,25} and iron deposition has been detected in the same SDGM structures where the volume loss also oc-

curred.²³ It is thought that excessive levels of iron in the brain may lead to tissue damage by the generation of reactive oxygen species, most probably through the Fenton reaction.^{26,27} In this process, ferrous iron (Fe^{2+}) donates an electron to hydrogen peroxide, with the resulting toxic products being ferric iron (Fe^{3+}), hydroxyl anion (OH^-), and the highly reactive hydroxyl radical (OH^\bullet).

Few studies have investigated iron deposition in the earliest stages of MS, such as in those with CIS.²⁸ Ceccarelli et al²⁹ reported increased T2 hypointensity in the head of the left caudate nucleus of patients with CIS compared with HC. However, a recent study using relaxometry did not show differences between patients with CIS and HC.¹⁹ Moreover, patients with RR-MS appear to show even higher iron content in the SDGM than patients with CIS, suggesting that disease progression may be related to pronounced iron deposition.^{18,19,30}

If increased levels of iron are causally related to SDGM damage and disease development, iron deposition must be present at the earliest stages of the disease, such as in patients with CIS.

This study used a SWI-filtered phase approach to characterize possible iron deposition in the SDGM of patients with CIS. Paramagnetic substances within the brain, mostly in the

Received October 31, 2011; accepted after revision November 23.

From the Buffalo Neuroimaging Analysis Center (J.H., N.B., M.H.-B., E.C., C.K., C.M., M.G.D., R.Z.) and The Jacobs Neurological Institute (B.W.-G., D.H., R.Z.), State University of New York at Buffalo, Buffalo, New York.

Please address correspondence to Robert Zivadinov, MD, PhD, FAAN, Department of Neurology, School of Medicine and Biomedical Sciences, Buffalo Neuroimaging Analysis Center, 100 High St., Buffalo, New York 14203; e-mail: rzivadinov@bnac.net



Indicates open access to non-subscribers at www.ajnr.org

<http://dx.doi.org/10.3174/ajnr.A3030>

form of ferritin, influence the frequency of proton spin and cause local magnetic field changes.³¹ The SWI-filtered phase imaging method uses a complex-space high-pass filter to retain only biologically relevant phase shifts, to generate a metric of iron attenuation.^{23,31}

In this study, we compared SDGM measurements of abnormal phase, indicative of increased iron content, and normalized volume between patients with CIS and a group of HC. Moreover, we sought to investigate the relationship of these measurements with conventional MR imaging techniques.

Materials and Methods

Subjects

We prospectively enrolled 42 consecutive patients with CIS (31 women, 11 men). Inclusion criteria were a diagnosis of CIS,³² age 18–65 years, and disease duration between 0 and 5 years. Sixty-five age- and sex-matched HC were also prospectively enrolled, all of whom had normal MR imaging and physical examinations. Patients with CIS underwent full neurologic assessment, including determination of EDSS.³³ Participants were excluded if they had a relapse or were treated with steroids within the month preceding study entry, were pregnant, or had any pre-existing medical conditions known to be associated with brain pathology. The internal institutional review board approved the study protocol and written informed consent was obtained from all participants.

Image Acquisition

A 3T Signa Excite HD 12.0 (GE Healthcare, Milwaukee, Wisconsin) with an 8-channel head and neck (HDNV) coil was used for scanning purposes. SWI data were acquired using a 3D flow-compensated gradient recalled-echo sequence with 64 locations, 2 mm thickness, a 512×192 (frequency \times phase) matrix, FOV = 25.6 cm \times 19.2 cm, with pFOV = 0.75, resulting in an in-plane resolution of 0.5 mm \times 1 mm. Other relevant parameters included flip angle = 12°, TE = 22, TR = 40 ms, and acquisition time = 8 min, 46 s.²³

In addition, we acquired the following sequences: 2D multiplanar dual FSE proton attenuation and T2-weighted imaging (TE1/TE2/TR = 9/98/5300 ms, flip angle = 90°, and ETL = 14; FLAIR (TE/TR = 120/8500 ms, TI = 2100 ms, flip angle = 90°, ETL = 24); 3D high-resolution T1WI using fast-spoiled gradient echo with magnetization-prepared inversion recovery pulse (TE/TI/TR = 2.8/900/5.9 ms, flip angle = 10°); and spin-echo T1WI (TE/TR = 16/600 ms, flip angle = 90°). All sequences except SWI were obtained with a 256×192 matrix, FOV = 25.6 cm \times 19.2 cm, pFOV = 0.75, for an in-plane resolution of 1 mm \times 1 mm. Forty-eight sections were collected, with a thickness of 3 mm, and no gap between sections, for all 2D scans (proton attenuation/T2, FLAIR, and spin-echo T1). For the 3D high-resolution inversion recovery fast-spoiled gradient echo, 184 1-mm-thick locations were acquired, resulting in isotropic resolution. All scans were prescribed in an axial-oblique orientation, parallel to the subcallosal line. One average was used for all sequences.

Image Analyses

Analyses were performed by operators who were unaware of the participants' disease status.

Abnormal Phase Identification

Raw (*k*-space) SWI data were transferred to an off-line Linux workstation for postprocessing using software developed in-house and

written in Matlab (MathWorks, Natick, Massachusetts). A detailed overview of SWI image acquisition, processing, analysis, and validation is provided elsewhere.²³

To segment SDGM structures, a combination of semiautomated edge-contouring and FMRIB's FIRST (<http://www.fmrib.ox.ac.uk/fsl/downloads/>) on 3D T1 WI was used.³⁴ Specifically, the thalamus, caudate, putamen, globus pallidus, hippocampus, amygdala, and nucleus accumbens were identified in this way.³⁵ Structures not identifiable by FIRST, such as the red nucleus, pulvinar nucleus of the thalamus, and substantia nigra, were identified semiautomatically using JIM5 (Xinapse Systems, Northamptonshire, United Kingdom) on the most representative section for each subject.³⁵

Using the SWI-filtered phase images, additional processing was performed to identify voxels likely to contain abnormal amounts of iron, based on their phase values. Reference phase values (means and SD) for each SDGM structure were determined previously using a large sample of HC.²³ Only the voxels of SDGM structures with phase values lower than 2 standard deviations below the reference group were retained. This yielded structure-specific maps of voxels with abnormally low phase. Subsequently, as a measure of the level of phase decrease, the mean value of subthreshold voxels was calculated to yield the MP-APT. More negative MP-APT values indicate increased iron content within that region, and mean values are reported in radians. In addition, we determined the SDGM NPTV by subtracting the abnormal phase tissue volume of a particular structure from the total volume of the same structure.

Global Atrophy and Lesion Analyses

The SIENAX cross-sectional software tool (version 2.6; <http://www.fmrib.ox.ac.uk/fsl/feeds/doc/index.html>) was used for brain extraction and tissue segmentation, with correction for T1-hypointensity misclassification.³⁶ As described previously,³⁷ we acquired the following volume measures: NBV, NGMV, NWMV, and NLVV. T2- and Gd-LVs and numbers were measured using a semiautomated edge detection contouring/thresholding technique, as described previously.³⁸ Normalized volumes were obtained for all SDGM structures using FIRST.³⁵

Statistical Analysis

Analyses were conducted using PASW Statistics, version 18.0 (IBM, Somers, New York). Differences between the groups were tested using the χ^2 -test (sex) and Student *t* test. Distributions of the data were tested for normality using the Shapiro-Wilk test. Because SDGM MP-APT, MP-APT volume, and NPTV were not normally distributed ($P < .001$), the Mann-Whitney *U* test was used to evaluate group differences of these variables. SDGM volume measurements were compared using the Student *t* test. Effect sizes are stated as Cohen *d*. We used Spearman correlation coefficients to assess the relationship between SDGM MP-APT, MP-APT volume, NPTV, and normalized volume with other MR imaging variables. Nominal *P* values $< .05$ were regarded as significant, using 2-tailed testing, and *P* values $< .10$ were considered a trend.

Results

Table 1 shows the demographic, clinical, and MR imaging characteristics of patients with CIS and HC. The mean age of patients with CIS was 40.1 years (SD = 10.4), with a mean disease duration of 1.2 years (SD = 1.3) and median EDSS score of 1.0 (range = 0–2.5). Thirty-one (73.8%) patients with CIS were female. Of the 42 patients with CIS, 22 (52.4%) were

Table 1: Demographic, clinical, and MRI characteristics of the study groups

	HC (<i>n</i> = 65)	CIS (<i>n</i> = 42)	<i>P</i> value
Female (<i>n</i> [%])	41 (63.1)	31 (73.8)	.295
Age in years (mean [SD])	42.8 (14)	40.1 (10.4)	.288
Age at onset in years (mean [SD])	NA	38.8 (10.6)	NA
Disease duration in years (mean [SD] median)	NA	1.2 (1.3) 1	NA
EDSS (median [range])	NA	1.0 (0–2.5)	NA
Presence of disease-modifying therapy	NA	22 (52.4%)	NA
Type of onset (<i>n</i> [%])			
Polysymptomatic	NA	7 (16.7%)	
Monosymptomatic	NA	35 (83.3%)	
Spinal cord syndrome		23 (65.7%)	
Optic neuritis		8 (22.9%)	
Brain stem/cerebellar		4 (11.4%)	
Gd lesion number (mean [SD])	NA	.4 (1.1)	NA
T2 lesion number (mean [SD])	NA	12.3 (15.3)	NA
1–4 lesions (<i>n</i> [%])		15 (35.7%)	
5–8 lesions (<i>n</i> [%])		6 (14.3%)	
≥9 lesions (<i>n</i> [%])		18 (42.9%)	
Gd lesion volume (mean [SD])	NA	.08 (.3)	NA
T2 lesion volume (mean [SD])	NA	1.7 (2.6)	NA
NGMV (mean [SD])	822.3 (63.2)	821.5 (58.2)	.953
NWMV (mean [SD])	775.3 (63.3)	760.6 (40.2)	.154
NBV (mean [SD])	1592.9 (99.6)	1583 (56.5)	.521
NLVV (mean [SD])	27.9 (13.7)	32.6 (18.9)	.145

Note:—All lesion and brain volumes are expressed in milliliters. Of the 42 patients with CIS, 33 presented with brain T2 lesions and 9 with spinal cord T2 lesions. Differences between the groups were tested using the χ^2 (gender) test and Student *t* test. NA indicates not available.

Table 2: Differences in the SDGM MP-APT between patients with CIS and HC

	MP-APT			%	<i>P</i>
	HC	CIS	<i>d</i>	Difference	Value
Total SDGM	−.143 (.019)	−.15 (.024)	.31	5	.171
Caudate	−.165 (.024)	−.171 (.013)	.31	3.6	.140
Putamen	−.169 (.036)	−.19 (.045)	.5	12.4	.029
Globus pallidus	−.177 (.04)	−.174 (.02)	.11	1.7	.652
Thalamus	−.09 (.017)	−.096 (.012)	.37	6.7	.067
Hippocampus	−.158 (.046)	−.151 (.037)	.16	4.4	.329
Amygdala	−.2 (.070)	−.209 (.074)	.14	4.5	.632
Nucleus accumbens	−.72 (.187)	−.794 (.253)	.33	10.3	.581
Red nucleus	−.228 (.036)	−.235 (.024)	.22	3.1	.422
Substantia nigra	−.303 (.056)	−.313 (.042)	.22	3.3	.381
Pulvinar	−.135 (.022)	−.148 (.019)	.61	9.6	.004

Note:—MP-APT is expressed in radians. Differences between groups were tested using Mann-Whitney *U* test.

on disease-modifying therapy and 35 (83.3%) had a monosymptomatic onset. No significant differences were observed in age, sex, or atrophy measures between patients with CIS and HC. Of the 42 patients with CIS, 33 presented with brain T2 lesions and 9 with spinal cord T2 lesions. Of those patients with CIS who had brain lesions, 18 (54.5%) showed ≥9 T2 hyperintense lesions, and 4 patients showed Gd-active lesions.

Table 2 displays MP-APT differences in SDGM structures between HC and patients with CIS. Both the pulvinar nucleus of the thalamus ($P = .004$) and putamen ($P = .029$) showed increased MP-APT, and there was a trend for increased MP-APT in the thalamus ($P = .067$). These SDGM structures showed an increase of 9.6%, 12.4%, and 6.7% in MP-APT, respectively.

Table 3: Differences in MP-APT volume between patients with CIS and HC

	MP-APT Volume			%	<i>P</i>
	HC	CIS	<i>d</i>	Difference	Value
Total SDGM	14.33 (2.07)	15.21 (1.96)	.44	6.1	.048
Caudate	2.33 (.54)	2.6 (.59)	.5	11.6	.035
Putamen	2.64 (.8)	3.1 (.79)	.58	17.4	.004
Globus pallidus	1.33 (.25)	1.37 (.29)	.16	3	.396
Thalamus	6.43 (.84)	6.51 (.87)	.09	1.2	.721
Hippocampus	1.15 (.59)	1.12 (.48)	.04	2.6	.861
Amygdala	.37 (.2)	.41 (.21)	.23	10.8	.118
Nucleus accumbens	.08 (.09)	.08 (.11)	.02	2.9	.630
Red nucleus	.02 (.02)	.03 (.02)	.18	15.2	.255
Substantia nigra	.07 (.03)	.07 (.03)	.12	5.9	.595
Pulvinar	.09 (.05)	.11 (.04)	.4	19.6	.045

Note:—All volumes are reported in milliliters. Differences between groups were tested using the Mann-Whitney *U* test.

Table 4: Differences in the SDGM normalized volumes between patients with CIS and HC

	Volume			%	<i>P</i>
	HC	CIS	<i>d</i>	Difference	Value
Total SDGM	45.22 (4.59)	45.43 (4.1)	.05	.5	.818
Caudate	6.75 (.99)	6.89 (.89)	.16	2.1	.437
Putamen	9.59 (1.22)	9.7 (1.1)	.1	1.1	.617
Globus pallidus	3.52 (.4)	3.44 (.35)	.2	−2.3	.329
Thalamus	15.21 (1.54)	15 (1.53)	.12	−1.4	.550
Hippocampus	7 (.86)	7.16 (.74)	.2	2.3	.324
Amygdala	2.38 (.36)	2.38 (.35)	.01	0	.963
Nucleus accumbens	.79 (.18)	.82 (.17)	.18	3.8	.400
Red nucleus	.17 (.02)	.17 (.03)	.04	0	.870
Substantia nigra	.29 (.05)	.32 (.05)	.6	10.3	.003
Pulvinar	.43 (.07)	.44 (.07)	.09	2.3	.619

Note:—All volumes are reported in milliliters. Differences between groups were tested using the Student *t* test.

Table 5: Differences in the NPTV between patients with CIS and HC

	NPTV			%	<i>P</i>
	HC	CIS	<i>d</i>	Difference	Value
Total SDGM	30.89 (3.56)	30.12 (3.33)	.2	−2.5	.203
Caudate	4.42 (.85)	4.29 (.81)	.15	−2.9	.558
Putamen	6.95 (1.25)	6.6 (1.26)	.27	−5	.189
Globus pallidus	2.2 (.33)	2.08 (.36)	.34	−5.5	.099
Thalamus	8.77 (.97)	8.51 (.93)	.27	−3	.120
Hippocampus	5.85 (.88)	6.04 (.8)	.22	3.2	.309
Amygdala	2 (.37)	1.97 (.39)	.11	−1.5	.415
Nucleus accumbens	.71 (.19)	.73 (.19)	.15	2.8	.623
Red nucleus	.15 (.03)	.14 (.03)	.08	−6.7	.346
Substantia nigra	.23 (.06)	.26 (.06)	.41	13	.048
Pulvinar	.33 (.07)	.32 (.09)	.14	−3	.356

Note:—All volumes are reported in milliliters. Differences between groups were tested using the Mann-Whitney *U* test.

Table 3 shows differences in MP-APT volume between patients with CIS and HC. A significantly increased volume of the MP-APT was observed in patients with CIS compared with HC in the putamen ($P = .004$), caudate nucleus ($P = .035$), pulvinar nucleus of the thalamus ($P = .045$), and total SDGM ($P = .048$).

Tables 4 and 5 show normalized volume and NPTV differences between patients with CIS and HC. No significant differences between the study groups were detected. The substantia nigra was significantly larger among patients with

CIS, compared with HC ($P = .003$), and had higher NPTV ($P = .048$).

To investigate the differences in patients with CIS, they were further divided into subgroups according to median T2-LV, T2 lesion number (≥ 9 and < 9), polysymptomatic versus monosymptomatic onset, different subtypes of monosymptomatic onset, and therapy status. No significant differences were found for any of these comparisons.

Increased MP-APT volume of the thalamus was related to increased T2-LV ($P = .033$). Furthermore, significant correlations were observed between the decreased NWMV measurements, with increased MP-APT volume in total SDGM, caudate, thalamus, hippocampus, and substantia nigra ($P < .05$). Decreased SDGM volume measurements in total SDGM, caudate nucleus, putamen, globus pallidus, thalamus, hippocampus, and red nucleus volume yielded significant relationships with decreased NWMV ($P < .01$). Moreover, increased T2-LV was inversely related to the decreased volume of the thalamus and the pulvinar nucleus of the thalamus ($P < .05$). Decreased NPTV of total SDGM, thalamus, the pulvinar nucleus of the thalamus, and hippocampus was also strongly related to decreased NWMV ($P < .01$).

No significant relationship was found between increased MP-APT or MP-APT volumes and decreased normalized volumes in the same individual SDGM structures.

Discussion

In this study, we investigated the extent of iron deposition using SWI-filtered phase measurements in the SDGM of patients with CIS and HC. We observed increased abnormal phase and abnormal phase volume in the SDGM of patients with CIS, compared with age- and sex-matched HC. Significant increases were observed in the total SDGM, caudate nucleus, putamen, and pulvinar nucleus of the thalamus. However, no differences were detected in volumetric measures of the same regions between the 2 study groups. These findings suggest that increased iron deposition in the SDGM structures may precede structure-specific atrophy development.

Iron deposition in the brain parenchyma of patients with MS has been observed using histopathologic techniques.^{24,25} More recently, MR imaging techniques, such as T2 relaxometry,¹⁷⁻¹⁹ T2 hypointensity,^{14-16,30,39} magnetic field correlation,²⁰ and SWI^{21-23,40} have been used to quantify and visualize iron deposition in vivo. Looking at changes in T2 intensity is a relatively straightforward method of evaluating iron content, because increased levels of iron result in reduced T2 relaxation time, leading to hypointensity on T2-weighted images.⁴¹ In MS, SDGM T2 hypointensity was found to be a predictor of clinical progression,³⁰ ambulatory impairment,¹⁶ cognitive impairment,^{15,19} and brain atrophy.^{14,19,42,43} SDGM T2 hypointensity has also been observed in patients with benign MS.⁴⁴ There is increasing evidence that iron deposition is also present in early stages of MS, such as in patients with pediatric MS and CIS.^{29,45} Ceccarelli et al²⁹ reported T2 hypointensity in the head of the left caudate nucleus of patients with CIS compared with HC. Moreover, atrophy of SDGM structures has also been observed in patients with CIS.³⁻¹⁰ Using R2* relaxometry, Khalil et al¹⁸ found significantly increased relaxation rates, indicative of iron deposition, in the caudate nucleus, putamen, and globus pallidus of patients

with RRMS compared with patients with CIS. In their most recent work,¹⁹ these authors reported that patients with CIS did not present with increased iron content compared with HC. However, the HC were approximately 3 years older than patients with CIS, who presented with much larger variance in age range compared with HC. The approach used in that study did not eliminate all those voxels classified as iron that were expected to be present in healthy subjects in relation to their aging process, with the highest content being in the globus pallidus, putamen, and caudate.²³ Differences in findings between our study and the latter one on iron content could be related to the sample size (we had twice the number of HC with lower age variance), use of different iron-based imaging techniques (SWI phase versus relaxometry), or detection of only abnormal iron content, when corrected for age. In contrast to these recent findings,¹⁹ our results suggest that excessive iron deposition is already present in patients with CIS and increases with disease progression.^{30,39}

In this study, we used an SWI-filtered phase approach, which has been histopathologically validated.⁴⁶ SWI is an MR imaging technique capable of visualizing tissues affected by iron deposition in the form of ferritin, deoxyhemoglobin, or hemosiderin.^{31,47} This method was recently used in a study finding strong iron content differences between patients with MS and HC in SDGM.²³ An SWI-filtered phase approach has several methodologic advantages over T2 hypointensity and relaxometry measurements, being more sensitive to detect tissue changes indicative of iron.^{23,31,47}

Iron deposition may be derived from myelin and oligodendrocyte debris, as well as iron stores in macrophages, or it may be the product of hemorrhages from damaged brain vessels.⁴⁸ Excessive iron may lead to the formation of hydroxyl radicals through Fenton and Haber-Weiss reactions, causing tissue damage.²⁷ The measurement of MP-APT allows for the quantification of only the phase values lower than 2 standard deviations below the mean of a previously defined representative control group.²³ Furthermore, we defined NPTV as brain tissue free of abnormal phase, which may reflect a healthier state of tissue. NPTV was derived by subtracting the abnormal phase tissue volume from the total volume of that same SDGM structure. Recently, robust differences were observed in NPTV of the SDGM using an adult sample of patients with MS compared with HC.²³ In this study, only a trend for a decrease of NPTV was observed in the globus pallidus. The significant increase in NPTV of the substantia nigra may be a result of increased statistical power to detect significance because this SDGM structure is known to have intrinsically high iron content.⁴⁹

It was recently reported that iron deposition is dominant in the pulvinar nucleus of the thalamus in a sample of adult patients with MS aged 19–35 years.⁵⁰ In addition to changes in the basal ganglia, we also found significant increases of pulvinar MP-APT and MP-APT volume, suggesting that this structure may be exceptionally prone to iron accumulation in MS. Because iron accumulation is observed in the pulvinar nucleus of the thalamus in patients with CIS, it stands to reason that it may be one of the earliest affected SDGM structures.

An important finding of our study was that, with exception of the substantia nigra, no SDGM volume loss was observed in patients with CIS. Moreover, global and region-specific vol-

ume measurements were also similar between the 2 study groups. In contrast, significant increases of MP-APT and MP-APT volume were observed in the SDGM of patients with CIS. Until now, it has been unclear whether iron deposition is merely an epiphenomenon of MS pathology or if it is an instigator of inflammation and disease development.^{19,51} In a recent study, Williams et al investigated the relationship between inflammation and iron deposition using an original animal model labeled as “cerebral experimental autoimmune encephalomyelitis,” which develops CNS perivascular iron deposits.⁵² They reported that inflammatory cell infiltrates were associated with perivascular iron deposits; however, inflammatory cells were also observed without associated iron deposits. There was an association between T2-LV accumulation and increased MP-APT volume and decreased normalized volumes and NPTV in various SDGM structures, especially the thalamus and pulvinar nucleus of thalamus. Therefore, this study provides novel information indicating that iron deposition and atrophy development in basal ganglia are closely related to inflammation, as measured by T2 and Gd lesion accumulation.

The findings from this study also suggest that iron deposition may occur before detectable atrophy in the same structures. In fact, no significant relationship was found between increased MP-APT or MP-APT volumes and decreased normalized volumes in the individual SDGM structures. A previous study showed a direct relationship between increased MP-APT and structure-specific atrophy in different SDGM regions in the later stages of the disease.²³ Therefore, future longitudinal studies should explore the interplay between iron deposition and atrophy development in individual SDGM structures from the first clinical onset.

Increased MP-APT, MP-APT volume, and decreased normalized volumes and NPTV in total SDGM, caudate, thalamus, pulvinar nucleus of the thalamus, putamen, globus pallidus, hippocampus, substantia nigra, and red nucleus were related with decreased NWMV but not with NGMV. These results indicate that there is a close relationship between iron deposition and WM, but not cortical GM atrophy development at the earliest stages of MS. These findings indicate that global GM atrophy may be preceded by inflammation and iron accumulation that is confined to SDGM and WM regions.

There are several potential limitations inherent in SWI-filtered phase imaging. Phase changes could be the result of changes to the underlying tissue structure or local iron distribution rather than absolute changes in iron content, and different chemical forms of iron could potentially have different SWI-phase effects.⁵³ Patients with CIS were relatively older and had somewhat longer disease duration than those enrolled in typical clinical trials.⁵⁴ Presumably, this was because some of the patients with CIS in our study were prospectively enrolled at the time of their referral to our MS specialty center. This could explain the somewhat higher disease duration than reported in clinical trials.⁵⁴ However, clinical and MR imaging features of the enrolled sample of patients with CIS are representative because the observed number and volume of T2 lesions and clinical disability are similar to previously reported baseline CIS cohorts.⁵⁴ In addition, the patients with CIS were approximately 2 years younger than HC. Nevertheless, future

studies should investigate iron deposition in younger patients with CIS.

However, we believe that by using a large sample, and data both on abnormal phase and atrophy measures, our study provides support for the concept that iron deposition is present at the earliest MS disease stages and may contribute to disease development and brain damage. As has been shown previously in patients with MS,²³ the pulvinar nucleus of the thalamus, as well as the basal ganglia, appear to exhibit the most widespread iron deposition in patients with CIS. Future prospective studies should also explore the relationship between the rate of iron deposition and conversion to clinically definite MS and development of disability in patients with CIS.

Disclosures: Bianca Weinstock-Guttman received personal compensation for consulting, speaking, and serving on a scientific advisory board for Biogen Idec, Teva Neuroscience, and EMD Serono. Dr. Weinstock-Guttman also received financial support for research activities from NMSS, NIH, ITN, Teva Neuroscience, Biogen Idec, EMD Serono, and Aspreva. **Additional ICMJE Disclosures:** UNRELATED: Consultancy: Pfizer, Acorda, Questcor; Grants/Grants Pending: ITN, Novartis, Shire; Payment for Lectures (including service on speakers bureaus): Biogen Idec, Teva Neuroscience, EMD Serono, Pfizer, Acorda.) David Hojnacki has received speaker honoraria and consultant fees from Biogen Idec, Teva Pharmaceutical Industries Ltd., EMD Serono, Pfizer Inc, and Novartis. Robert Zivadinov received personal compensation from Teva Neuroscience, Biogen Idec, EMD Serono and Questcor Pharmaceuticals for speaking and consultant fees. Dr. Zivadinov received financial support for research activities from Biogen Idec, Teva Neuroscience, Genzyme, Bracco, Questcor Pharmaceuticals and EMD Serono.

Acknowledgments

The authors thank Eve Salczynski for technical assistance in the preparation of this manuscript.

References

- Pirko I, Lucchinetti CF, Sriram S, et al. **Gray matter involvement in multiple sclerosis.** *Neurology* 2007;68:634–42
- Fisher E, Lee JC, Nakamura K, et al. **Gray matter atrophy in multiple sclerosis: a longitudinal study.** *Ann Neurol* 2008;64:255–65
- Audoin B, Ibarrola D, Malikova I, et al. **Onset and underpinnings of white matter atrophy at the very early stage of multiple sclerosis—a two-year longitudinal MRI/MRSI study of corpus callosum.** *Mult Scler* 2007;13:41–51
- Audoin B, Zaaraoui W, Reuter F, et al. **Atrophy mainly affects the limbic system and the deep grey matter at the first stage of multiple sclerosis.** *J Neurol Neurosurg Psychiatry* 2010;81:690–95
- Calabrese M, Atzori M, Bernardi V, et al. **Cortical atrophy is relevant in multiple sclerosis at clinical onset.** *J Neurol* 2007;254:1212–20
- Henry RG, Shieh M, Amirbekian B, et al. **Connecting white matter injury and thalamic atrophy in clinically isolated syndromes.** *J Neurol Sci* 2009;282:61–66
- Henry RG, Shieh M, Okuda DT, et al. **Regional grey matter atrophy in clinically isolated syndromes at presentation.** *J Neurol Neurosurg Psychiatry* 2008;79:1236–44
- Ramasamy DP, Benedict RH, Cox JL, et al. **Extent of cerebellum, subcortical and cortical atrophy in patients with MS: a case-control study.** *J Neurol Sci* 2009;282:47–54
- Roosendaal SD, Bendfeldt K, Vrenken H, et al. **Grey matter volume in a large cohort of MS patients: relation to MRI parameters and disability.** *Mult Scler* 2011;17:1098–106
- Fisniku LK, Chard DT, Jackson JS, et al. **Gray matter atrophy is related to long-term disability in multiple sclerosis.** *Ann Neurol* 2008;64:247–54
- Calabrese M, Rinaldi F, Mattisi I, et al. **The predictive value of gray matter atrophy in clinically isolated syndromes.** *Neurology* 2011;77:257–63
- Dalton CM, Chard DT, Davies GR, et al. **Early development of multiple sclerosis is associated with progressive grey matter atrophy in patients presenting with clinically isolated syndromes.** *Brain* 2004;127:1101–07
- Raz E, Cercignani M, Sbardella E, et al. **Gray- and white-matter changes 1 year after first clinical episode of multiple sclerosis: MR imaging.** *Radiology* 2010;257:448–54
- Bakshi R, Benedict RHB, Bermel RA, et al. **T2 hypointensity in the deep gray matter of patients with multiple sclerosis: a quantitative magnetic resonance imaging study.** *Arch Neurol* 2002;59:62
- Brass SD, Benedict RHB, Weinstock-Guttman B, et al. **Cognitive impairment is**

- associated with subcortical magnetic resonance imaging grey matter T2 hypointensity in multiple sclerosis. *Mult Scler* 2006;12:437
16. Tjoa CW, Benedict RH, Weinstock-Guttman B, et al. MRI T2 hypointensity of the dentate nucleus is related to ambulatory impairment in multiple sclerosis. *J Neurol Sci* 2005;234:17–24
 17. Burgetova A, Seidl Z, Krasensky J, et al. Multiple sclerosis and the accumulation of iron in the basal ganglia: quantitative assessment of brain iron using MRI T(2) relaxometry. *Eur Neurol* 2010;63:136–43
 18. Khalil M, Enzinger C, Langkammer C, et al. Quantitative assessment of brain iron by R(2)* relaxometry in patients with clinically isolated syndrome and relapsing-remitting multiple sclerosis. *Mult Scler* 2009;15:1048–54
 19. Khalil M, Langkammer C, Ropele S, et al. Determinants of brain iron in multiple sclerosis: A quantitative 3T MRI study. *Neurology* 2011;77:1691–97
 20. Ge Y, Jensen JH, Lu H, et al. Quantitative assessment of iron accumulation in the deep gray matter of multiple sclerosis by magnetic field correlation imaging. *AJNR Am J Neuroradiol* 2007;28:1639
 21. Eissa A, Lebel RM, Korzan JR, et al. Detecting lesions in multiple sclerosis at 4.7 Tesla using phase susceptibility-weighting and T2-weighting. *J Magn Reson Imaging* 2009;30:737–42
 22. Haacke EM, Makki M, Ge Y, et al. Characterizing iron deposition in multiple sclerosis lesions using susceptibility weighted imaging. *J Magn Reson Imaging* 2009;29:537–44
 23. Zivadinov R, Heininen-Brown M, Schirda C, et al. Abnormal subcortical deep-gray matter susceptibility-weighted imaging filtered phase measurements in patients with multiple sclerosis. A case-control study. *Neuroimage* 2012;59:331–39
 24. Craelius W, Migdal MW, Luessenhop CP, et al. Iron deposits surrounding multiple sclerosis plaques. *Arch Pathol Lab Med* 1982;106:397–99
 25. LeVine SM. Iron deposits in multiple sclerosis and Alzheimer's disease brains. *Brain Res* 1997;760:298–303
 26. Fenton HJH. Oxidation of tartaric acid in presence of iron. *J Chem Soc* 1894;65:899–903
 27. Khalil M, Teunissen C, and Langkammer C. Iron and neurodegeneration in multiple sclerosis. *Mult Scler Int* 2011;606807
 28. Miller D, Barkhof F, Montalban X, et al. Clinically isolated syndromes suggestive of multiple sclerosis, part I: natural history, pathogenesis, diagnosis, and prognosis. *Lancet Neurol* 2005;4:281–88
 29. Ceccarelli A, Rocca MA, Neema M, et al. Deep gray matter T2 hypointensity is present in patients with clinically isolated syndromes suggestive of multiple sclerosis. *Mult Scler* 2010;16:39–44
 30. Neema M, Arora A, Healy BC, et al. Deep gray matter involvement on brain MRI scans is associated with clinical progression in multiple sclerosis. *J Neuroimaging* 2009;19:3–8
 31. Haacke EM, Mittal S, Wu Z, et al. Susceptibility-weighted imaging: technical aspects and clinical applications, part 1. *AJNR Am J Neuroradiol* 2009;30:19
 32. Polman CH, Reingold SC, Banwell B, et al. Diagnostic criteria for multiple sclerosis: 2010 revisions to the McDonald criteria. *Ann Neurol* 2011;69:292–302
 33. Kurtzke JF. Rating neurologic impairment in multiple sclerosis: an Expanded Disability Status Scale (EDSS). *Neurology* 1983;33:1444–52
 34. Patenaude B, Smith SM, Kennedy DN, et al. A Bayesian model of shape and appearance for subcortical brain segmentation. *Neuroimage* 2011;56:907–22
 35. Batista S, Zivadinov R, Hoogs M, et al. Basal ganglia, thalamus and neocortical atrophy predicting slowed cognitive processing in multiple sclerosis. *J Neurol* 2012;259:139–46
 36. Yeh EA, Weinstock-Guttman B, Ramanathan M, et al. Magnetic resonance imaging characteristics of children and adults with paediatric-onset multiple sclerosis. *Brain* 2009;132:3392–400
 37. Zivadinov R, Weinstock-Guttman B, Benedict R, et al. Preservation of gray matter volume in multiple sclerosis patients with the Met allele of the rs6265 (Val66Met) SNP of brain-derived neurotrophic factor. *Hum Mol Genet* 2007;16:2659–68
 38. Zivadinov R, Rudick RA, De Masi R, et al. Effects of IV methylprednisolone on brain atrophy in relapsing-remitting MS. *Neurology* 2001;57:1239–47
 39. Bakshi R, Shaikh ZA, Janardhan V. MRI T2 shortening (“black T2”) in multiple sclerosis: frequency, location, and clinical correlation. *Neuroreport* 2000;11:15–21
 40. Hammond KE, Metcalf M, Carvajal L, et al. Quantitative in vivo magnetic resonance imaging of multiple sclerosis at 7 Tesla with sensitivity to iron. *Ann Neurol* 2008;64:707–13
 41. Drayer B, Burger P, Darwin R, et al. MRI of brain iron. *AJR Am J Roentgenol* 1986;147:103–10
 42. Bakshi R, Dmochowski J, Shaikh ZA, et al. Gray matter T2 hypointensity is related to plaques and atrophy in the brains of multiple sclerosis patients. *J Neurol Sci* 2001;185:19–26
 43. Bermel RA, Puli SR, Rudick RA, et al. Prediction of longitudinal brain atrophy in multiple sclerosis by gray matter magnetic resonance imaging T2 hypointensity. *Arch Neurol* 2005;62:1371–76
 44. Ceccarelli A, Filippi M, Neema M, et al. T2 hypointensity in the deep gray matter of patients with benign multiple sclerosis. *Mult Scler* 2009;15:678–86
 45. Ceccarelli A, Rocca MA, Perego E, et al. Deep grey matter T2 hypo-intensity in patients with paediatric multiple sclerosis. *Mult Scler* 2011;17:702–07
 46. Hopp K, Popescu BF, McCrea RP, et al. Brain iron detected by SWI high pass filtered phase calibrated with synchrotron X-ray fluorescence. *J Magn Reson Imaging* 2010;31:1346–54
 47. Haacke EM, Xu Y, Cheng YC, et al. Susceptibility weighted imaging (SWI). *Magn Reson Med* 2004;52:612–18
 48. Stankiewicz J, Panter SS, Neema M, et al. Iron in chronic brain disorders: imaging and neurotherapeutic implications. *Neurotherapeutics* 2007;4:371–86
 49. Snyder AM, Connor JR. Iron, the substantia nigra and related neurological disorders. *Biochim Biophys Acta* 2009;1790:606–14
 50. Haacke EM, Garbern J, Miao Y, et al. Iron stores and cerebral veins in MS studied by susceptibility weighted imaging. *Int Angiol* 2010;29:149–57
 51. Zivadinov R, Weinstock-Guttman B, Pirkio I. Iron deposition and inflammation in multiple sclerosis. Which one comes first? *BMC Neurosci* 2011;12:60
 52. Williams R, Rohr AM, Wang WT, et al. Iron deposition is independent of cellular inflammation in a cerebral model of multiple sclerosis. *BMC Neurosci* 2011;12:59
 53. Schweser F, Deistung A, Lehr BW, et al. Quantitative imaging of intrinsic magnetic tissue properties using MRI signal phase: An approach to in vivo brain iron metabolism? *Neuroimage* 2011;54:2789–807
 54. Kappos L, Freedman MS, Polman CH, et al. Long-term effect of early treatment with interferon beta-1b after a first clinical event suggestive of multiple sclerosis: 5-year active treatment extension of the phase 3 BENEFIT trial. *Lancet Neurol* 2009;8:987–97