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Identification of *BRAF p. V600E*-Mutant and Wild-Type by MR Imaging in Pleomorphic Xanthoastrocytoma and Anaplastic Pleomorphic Xanthoastrocytoma

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ABSTRACT

BACKGROUND AND PURPOSE: Compared with *BRAF p. V600E* wild-type pleomorphic xanthoastrocytoma, *BRAF p. V600E*-mutant pleomorphic xanthoastrocytoma showed a higher survival rate. In this study, we focused on finding preoperative MR imaging differences between *BRAF p. V600E* mutant and wild-type in pleomorphic xanthoastrocytoma and anaplastic pleomorphic xanthoastrocytoma.

MATERIALS AND METHODS: Twenty-three patients with pathologically confirmed pleomorphic xanthoastrocytoma or anaplastic pleomorphic xanthoastrocytoma in our hospital were retrospectively analyzed from January 2015 to December 2020. They were divided into a *BRAF p. V600E*-mutant group (including 6 pleomorphic xanthoastrocytomas and 5 anaplastic pleomorphic xanthoastrocytomas) and a wild-type group (including 8 pleomorphic xanthoastrocytomas and 4 anaplastic pleomorphic xanthoastrocytomas). The preoperative MR imaging characteristics of these groups were statistically compared.

RESULTS: The wild-type pleomorphic xanthoastrocytoma group presented with more aggressive conventional and advanced MR imaging features than the mutant pleomorphic xanthoastrocytoma group, including greater mean maximum tumor diameter (3.1 [SD, 0.9] cm versus 1.7 [SD, 0.4] cm, $P < .05$), more frequent heterogeneous contrast enhancement of solid portions (100% versus 0%, $P < .001$), more obvious peritumoral edema (mean, [2.1 SD, 0.7] cm versus 0.6 [SD, 0.2] cm, $P < .01$), and lower mean minimum relative ADC (896 [SD, 86] versus 988 [SD, 73], $P < .05$) and mean relative ADC (1060 [SD, 159] versus 1248 [SD, 116], $P < .05$) on DWI. However, there was no significant difference in either conventional or advanced MR imaging features between the wild-type anaplastic pleomorphic xanthoastrocytoma group and the mutant anaplastic pleomorphic xanthoastrocytoma group.

CONCLUSIONS: Neurosurgeons should carefully interpret MR images before an operation and select appropriate surgical strategies according to genotype prediction.

ABBREVIATIONS: ADCmean = mean relative ADC; ADCmin = minimum relative ADC; APXA = anaplastic pleomorphic xanthoastrocytoma; PXA = pleomorphic xanthoastrocytoma; WHO = World Health Organization

Pleomorphic xanthoastrocytoma (PXA) is an infrequent glioma affecting 1% of patients with brain tumors, thought to originate from subpial astrocytes or their precursors and usually occurring in children and adolescents.¹ PXA was first described by Kepes et al,²

in 1979. In 1993, it was officially included in the World Health Organization (WHO) Classification System for Tumors of the Central Nervous System as a grade II tumor.³ Although PXA is classified as a WHO grade II tumor, “PXA with anaplastic features” comprise 15%–50% of these lesions.^{4–7} According to the 2016 WHO classification system, PXA is divided into 2 distinct entities based on histopathologic characteristics: WHO grade II PXA and WHO grade III anaplastic PXA (APXA).^{8,9} APXA is defined as the presence of 5 mitoses per 10 high-power fields. Patients with PXA can undergo only total resection, while patients with APXA require total resection and adjuvant therapy.^{5,10} In addition, APXA has been reported to have a worse prognosis than PXA, with a 5-year overall survival rate of 57.1%.^{5,11,12}

The most frequently found mutant gene in PXA is *BRAF p. V600E*, which encodes an intracellular component of the MAPK pathway.^{13–16} *BRAF p. V600E*-mutants were found in 70% of typical PXAs but appeared less commonly in APXA (17%–

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65%).^{5,13,17,18} *BRAF p. V600E*-mutant may divide these tumors into 2 clinically relevant subsets, both associated with natural history and response to treatment.¹⁹ Compared with the wild-type, *BRAF p. V600E*-mutant PXA showed a higher survival rate.^{5,20,21} *BRAF p. V600E*-mutant PXA has also shown a response to *BRAF*-targeted therapy,²² though no prospective comparative trials have been conducted in this patient population.

APXA can present with more aggressive conventional and advanced MR imaging features, mimicking high-grade astrocytoma at initial diagnosis, than PXA.²³ However, there have been few reports on the MR imaging characteristics of PXA with *BRAF p. V600E*-mutant, especially its differentiation from *BRAF p. V600E* wild-type PXA. Therefore, the purpose of this study was to investigate the MR imaging characteristics of *BRAF p. V600E*-mutant PXA and *BRAF p. V600E* wild-type PXAs to determine whether there are differences in the imaging characteristics of these 2 entities to aid in planning a treatment strategy and predicting prognosis.

MATERIALS AND METHODS

Case Selection

The study was approved by the Branch for Medical Research and Clinical Technology Application, Ethics Committee of the First Affiliated Hospital of Fujian Medical University and informed consent was signed. Of 317 astrocytic tumor cases from January 2015 to December 2020, twenty-eight (0.89%) with diagnoses containing the term “pleomorphic xanthoastrocytoma” or “anaplastic pleomorphic xanthoastrocytoma” on first surgical resection were extracted. Three (of 28) cases lacked MR imaging data before the first operation and were excluded. Twenty-five adult patients with PXA were included in this study, and a review of all their clinical and radiologic records was performed. The analyzed clinical information included sex, age, and symptoms (seizures, increased intracranial pressure, and neurologic deficits). These data were obtained from the electronic medical record system.

Histopathologic and Molecular Sequencing

Paraffin-embedded tumor tissue was cut into 4-mm sections and stained with H&E, and 10-mm sections were used for molecular sequencing. All histologic specimens were reviewed by our institutional neuropathologist (with 15 years of experience in neuropathology), who reclassified PXA (WHO grade II) and APXA (WHO grade III) according to the 2016 WHO classification of central nervous system tumors and the Consortium to Inform Molecular and Practical Approaches to CNS Tumor Taxonomy (cIMPACT-NOW) update 6.²⁴ The diagnostic criteria included tumors demonstrating a relatively solid growth pattern composed of spindle-shaped, pleomorphic, and multinucleated cells associated with both pale and bright eosinophilic granular bodies to be diagnosed as PXA. Anaplastic features, which included a mitotic index of 5 of 10 high-power fields with or without necrosis and endothelial proliferation, were diagnosed as APXA. Twenty-five patients were studied. Of these, 10 PXA and 13 APXA specimens were confirmed. We excluded 2 cases: One was reclassified as “diffuse astrocytoma, *IDH* wild-type with pleomorphic/PXA-like features,” and the other was previously diagnosed as “pleomorphic xanthoastrocytoma with anaplastic features” and reclassified as

“giant cell glioblastoma.” All 23 cases were available for molecular sequencing. Molecular analyses were performed to test the presence of the *BRAF p. V600E*-mutant, which was confirmed by sequencing. Of these, 11 *BRAF p. V600E*-mutant PXAs (including 6 cases of PXA and 5 cases of APXA) and 12 *BRAF p. V600E* wild-type PXAs (including 4 cases of PXA and 8 cases of APXA) were confirmed.

Preoperative MR Imaging Techniques

All patients underwent MR imaging on 3T scanners (Magnetom Verio or Magnetom Skyra; Siemens) or a 1.5T scanner (Signa TwinSpeed; GE Healthcare). The retrospective nature of this study resulted in variability in MR images and imaging protocols. Conventional MR imaging protocols included an axial T2-weighted sequence, nonenhanced axial and sagittal T1-weighted sequences, and 3 orthogonal plane contrast-enhanced T1-weighted sequences. An FOV of 22 cm², imaging matrix of 256 × 256, and section thickness of 5 mm were uniformly applied in all sequences.

DWI was performed in 10 patients with PXA and 13 patients with APXA using an axial echo-planar sequence according to the following technique parameters: TR/TE = 8200/102 ms, FOV = 22 × 22 cm, section thickness/gap = 5/1 mm, diffusion gradient encoding = $b = 0$, 1000 s/mm². The diffusion gradient was used in 3 orthogonal directions. ADC maps were automatically calculated using a monoexponential model.

MR Imaging Analysis

All images were reviewed in consensus by 2 radiologists (readers 1 and 2 with 20 and 8 years of experience in neuroimaging, respectively) to make a factual comparison and minimize the confounding effects. The readers were blinded to tumor histology and recorded the following tumor characteristics: 1) tumor location (frontal, occipital, temporal, or parietal; superficial or deep); 2) tumor size (largest diameter, in centimeters); 3) the presence of cystic degeneration; 4) enhancement characteristics of the solid component (heterogeneous or homogeneous); 5) the presence and degree of peritumoral edema (largest diameter, in centimeters); 6) leptomeningeal contact; and 7) the presence of restricted diffusion, defined as high signal on DWI and corresponding low signal on ADC maps compared with contralateral normal brain parenchyma.

Tumor size was defined by the maximum diameter as measured on contrast-enhanced T1WI. The extent of peritumoral edema was defined by a region of very bright signal intensity on T2 surrounding the enhanced tumor on contrast-enhanced T1WI. The tumor location was defined as main lobe involvement when >1 lobe was involved.

For quantitative ADC evaluation, MR images were postprocessed as follows: 1) identification of the axial DWI and corresponding ADC image section displaying the lowest ADC values within solid tumor tissue components; 2) definition of a circular ROI of 6-mm diameter, manually drawn within the perceived lowest ADC solid tumor component, recording the minimum relative ADC (ADC_{min}) and mean relative ADC (ADC_{mean}) values of this tumoral ROI (the ROI was systematically moved within the solid tumoral area, as depicted by conventional MR imaging, to measure regions with the lowest ADC values); 3)

Table 1: Demographic data and conventional MR imaging characteristics of PXA and APXA

	PXA	APXA	P Value
Clinical data			
Male sex (No.) (%)	3 (30%)	6 (46.1%)	.363
Mean age (yr)	34 (SD, 10)	41 (SD, 10)	.232
Presenting symptoms (No.)			.899
Seizures	6	9	
Headache or increased ICP	2	2	
Neurologic deficit	2	2	
<i>BRAF p. V600E</i> -mutant	6 (60%)	5 (38.5%)	.273
Location (No.)			.526
Frontal lobe	0	3	
Temporal lobe	5	5	
Occipital lobe	1	1	
Parietal lobe	2	3	
Insula	2	1	
Superficial location (No.) (%)	8 (80%)	11 (84.6%)	.596
Conventional MR imaging			
Mean size (cm)	2.2 (SD, 0.9)	3.2 (SD, 0.6)	<.01
Presence of cystic degeneration (No.) (%)	6 (60%)	9 (69.2%)	.490
Peritumoral edema (mean) (cm)	1.2 (SD, 0.8)	2.1 (SD, 0.9)	.021
Heterogeneous enhancement (No.) (%)	4 (40%)	11 (84.6%)	<.001
Leptomeningeal contact (No.) (%)	8 (80%)	11 (84.6%)	.596

Note:—ICP indicates intracranial pressure.

definition and assessment of the reference region minimum and mean ADC values, by placing a similar ROI on the normal-appearing thalamus (because it is a readily identifiable structure on ADC maps and is considered generally resistant to changes related to hydrocephalus);²⁵ and 4) finally, the mean and minimum ADC values of the tumoral ROIs with the lowest ADC value were normalized with their corresponding reference region, obtaining ADCmean and ADCmin. This computation was performed because ADC values of normal brain vary with age,²⁶ and the contrast achieved between tumor and surrounding tissue depends on this ratio; at the same time, this computation allowed data comparison of brain MR imaging performed with different scanners.

Statistical Analysis

The normality of all continuous parameters was initially assessed using the Kolmogorov-Smirnov test. Characteristics of the patients were summarized with means or medians and SDs or ranges for continuous data (as appropriate) and with frequencies and percentages for categoric data. The Fisher exact test was used to assess the differences in the categoric variables (age, sex, location, cystic degeneration, enhancement characteristics, and leptomeningeal contact) between the different groups. The Mann-Whitney *U* test was used to assess the differences in continuous variables (edema, tumor size, and ADC values) between the different groups because they did not conform to the normal distribution.

The MR imaging differences between the *BRAF p. V600E*-mutant PXA group and the *BRAF p. V600E* wild-type PXA group and the *BRAF p. V600E*-mutant APXA group and the *BRAF p. V600E* wild-type APXA group were statistically analyzed. Statistical analysis was performed with the Statistical Package for the Social Sciences (SPSS 24.0 Version for Windows; IBM). All reported *P* values were double-tailed, and *P* values less < .05 were regarded as statistically significant.

RESULTS

Clinical Data and Conventional MR Imaging

According to molecular analysis, 10 cases of PXA were divided into a *BRAF p. V600E*-mutant PXA group (*n* = 6) and a wild-type PXA group (*n* = 4); 13 cases of APXA were divided into a *BRAF p. V600E*-mutant APXA group (*n* = 5) and a wild-type APXA group (*n* = 8). Clinical data, molecular analysis, location, and conventional imaging features of PXA and APXA are summarized in Table 1. All 23 tumors were in the supratentorial region. Eight PXAs (8/10, 80%) and 11 APXAs (11/13, 84.6%) were superficial (on the surface of the brain parenchyma). There was no significant difference in tumor location between the PXA and APXA groups. The presence of heterogeneous enhancement of solid portions was observed less frequently in patients with PXA than in

patients with APXA (4/10 versus 11/13, *P* < .001). Leptomeningeal contact was seen in 8 PXA cases and 11 APXA cases. In preoperative images, the lesion was significantly larger in patients with APXA than in patients with PXA (mean, 3.2 [SD, 0.6] cm versus 2.2 [SD, 0.9] cm, *P* < .01). The average maximum diameter of peritumoral edema in the APXA group was larger than the average maximum diameter of peritumoral edema in the PXA group (2.1 [SD, 0.9] cm versus 1.2 [SD, 0.8] cm, *P* = .021) (Fig 1).

Table 2 and Table 3 summarize the clinical data, localization, and conventional imaging features of *BRAF p. V600E* subtypes of PXAs and APXAs, respectively. There was no significant difference in location between the *BRAF V600E* mutant and wild-type groups for either PXAs or APXAs. In preoperative images, the presence of heterogeneous enhancement of solid portions was observed less frequently in patients with *BRAF p. V600E*-mutant PXA than in patients with wild-type PXA (0/6 versus 4/4, *P* < .001). However, there was no significant difference in the presence of heterogeneous enhancement of the location of solid portions between the 2 groups in APXAs (5/5 versus 6/8, *P* = .359). The mean tumor size (the maximum diameter represents the size of the tumor) was significantly larger in patients with *BRAF p. V600E* wild-type PXA than in patients in the group with mutant PXA (3.1 [SD, 0.9] cm versus 1.7 [SD, 0.4] cm, *P* = .038). The maximum diameter of peritumoral edema in the *BRAF p. V600E* wild-type PXA group was significantly larger than the mutant PXA group (mean, 2.1 [SD, 0.7] cm versus 0.6 [SD, 0.2] cm, *P* < .01). However, there was no significant difference in either mean tumor size (3.3 [SD, 0.7] cm versus 3.2 [SD, 0.5] cm, *P* = .943) or the mean maximum diameter of peritumoral edema (2.4 [SD, 0.9] cm versus 1.8 [SD, 0.4] cm, *P* = .3) between the *BRAF p. V600E*-mutant type APXA group and the wild-type APXA group (Fig 2).

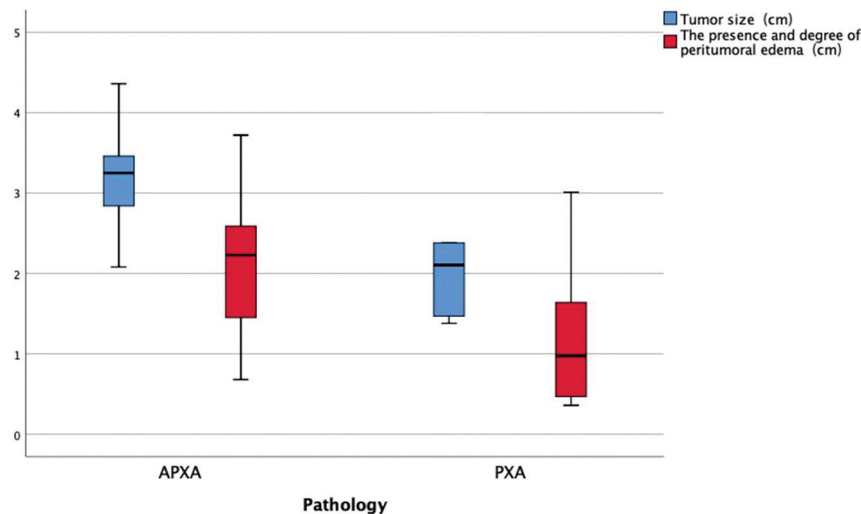


FIG 1. Boxplots representing tumor size and the presence and degree of peritumoral edema of PXA and APXA. A significant difference ($P < .05$) between PXA and APXA is demonstrated with both tumor size and the presence and degree of peritumoral edema.

Table 2: Demographic data and conventional MR imaging characteristics of *BRAF p. V600E*-mutant and *BRAF p. V600E* wild-type PXA

	<i>BRAF p. V600E</i> - Mutant PXA	<i>BRAF p. V600E</i> Wild- Type PXA	<i>P</i> Value
Clinical data			
Male sex (No.) (%)	2 (33.3%)	1 (25%)	.667
Mean age (yr)	32 (SD, 8.6)	37 (SD, 13)	.762
Presenting symptoms (No.)			.870
Seizures	4	2	
Headache or increased ICP	1	1	
Neurologic deficit	1	1	
Location (No.)			.405
Frontal lobe	0	0	
Temporal lobe	3	2	
Occipital lobe	0	1	
Parietal lobe	2	0	
Insula	1	1	
Superficial location (No.) (%)	4 (66.6%)	4 (100%)	.333
Conventional MR imaging			
Mean size (cm)	1.7 (SD, 0.4)	3.1 (SD, 0.9)	.038
Presence of cystic degeneration (No.) (%)	2 (33.3%)	4 (100%)	.071
Peritumoral edema (mean) (cm)	0.6 $\pm$ 0.2	2.1 $\pm$ 0.7	<.01
Heterogeneous enhancement (No.) (%)	0 (0%)	4 (100%)	<.001
Leptomeningeal contact (No.) (%)	6 (100%)	2 (50%)	.03

Note:— ICP indicates intracranial pressure.

Advanced MR Imaging Findings

Table 4 and Figure 3 summarize the features of PXAs and APXAs with DWI. Both mean ADCmin values (755 [SD, 77] versus 951 [SD, 88]; $P < .001$) and ADCmean values (956 [SD, 106] versus 1173 [SD, 159], $P < .001$) were significantly lower in the whole APXA group than in the PXA group. Both mean ADCmin values (896 [SD, 86] versus 988 [SD, 73], $P < .05$) and ADCmean values (1060 [SD, 159] versus 1248 [SD, 116], $P < .05$) were significantly lower in the whole group of *BRAF p. V600E* wild-type PXA than in mutant-PXA group. However, there was no significant difference in mean ADCmin (750 [SD, 73] versus 762 [SD, 92], $P = 1$) and

ADCmean (974 [SD, 92] versus 925 [SD, 131], $P = .724$) values between the 2 groups in APXAs (Fig 4, Tables 5 and 6). Representative cases are shown in Figure 5.

DISCUSSION

To the best of our knowledge, few studies have evaluated and compared the imaging features of *BRAF p. V600E* wild-type PXA and *BRAF p. V600E*-mutant PXA. Our results show that PXAs with *BRAF p. V600E* wild-type show more aggressive MR imaging features than those with the *BRAF p. V600E*-mutant according to MR imaging features, namely, larger tumor size, more heterogeneous contrast enhancement, obvious peritumoral edema, lower ADCmin, and lower ADCmean ratio on DWI.

PXA is histologically classified as a grade II astrocytoma according to the 2016 WHO classification of the central nervous system.⁸ The outcomes in cases of PXAs are relatively favorable,

with 5- and 10-year overall survival rates of 75%–81% and 67%–70%, respectively, after total resection.^{7,27} APXA is more likely to be derived from a previous PXA with the development of anaplastic histologic features.²⁸ In the present study, APXA accounted for 56.5% of PXAs and showed anaplastic features. Findings comparable with the results reported by Hirose et al⁶ were that approximately 50% of PXAs showed anaplastic features.

The *BRAF p. V600E*-mutant is a distinctive genomic alteration of PXA and APXA.^{17,29,30} Previous studies demonstrated that *BRAF p. V600E*-mutant status is an independent prognostic factor

after adjusting for anaplasia, age, sex, and initial symptoms.¹² In the present study, approximately 60% of PXAs and 38.4% of APXAs showed a *BRAF p. V600E*-mutant, findings comparable with the results reported by Dias-Santagata et al¹³ that *BRAF p. V600E*-mutants are found in 70% of typical PXAs but appear less commonly in APXAs.

Most large case series have reported no sex predilection. However, the proportion of women in our series was much higher than the proportion of men. PXA most frequently occurred in young adults in our study, consistent with previous studies.^{1,5,27} APXA was reported to be more common in older adults with a median age of 40–65 years.^{31–33} In our series, 9 of 13 patients with APXA were middle-aged adults with a mean age of

44.67 years at their first presentation, which is consistent with findings in previous studies.²⁷

The most common tumor location of PXA was the temporal lobe (50.0%), which is in accordance with findings in previous studies.^{34,35} Our results show that the principal initial manifestation was seizures, which were observed in 60% and 69.2% of patients with PXA and APXA, respectively. These results correspond to findings that seizures were obviously related to tumor involvement of the temporal lobe in PXA.¹² Almost all PXA and APXA tumors in our series showed a superficial location in the cerebral hemispheres with leptomeningeal contact. Both PXA and APXA are believed to develop from subpial astrocytes, which partly explains why these 2 tumors preferably arise from superficial cortical sites.²

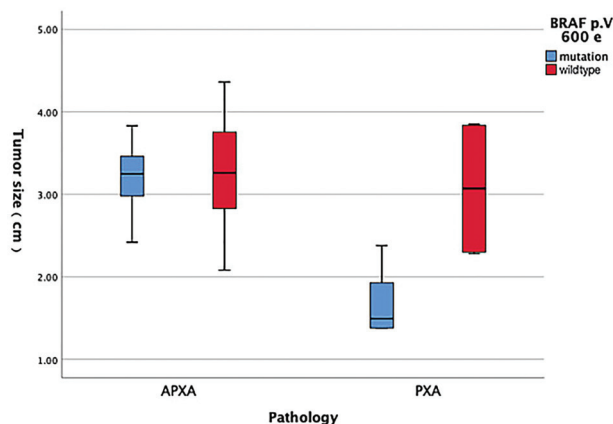
In our study, we found that APXA shows more aggressive MR imaging features than PXA, namely, larger tumor size, more heterogeneous contrast enhancement, and obvious peritumoral edema, consistent with previously reported findings.²³ *BRAF p. V600E*-mutant PXAs have been reported to show histologic heterogeneity and display pleomorphic giant cells, xanthomatous changes, and fascicular and storiform growth patterns.¹³ Purkait et al³⁶ also proposed the possibility of a morphologic spectrum of *BRAF p. V600E*-mutant tumors with astroblastoma at one end and PXA at the other. We found that *BRAF p. V600E* wild-type PXA showed significantly different imaging characteristics from mutant PXA, including larger tumor size, more heterogeneous contrast enhancement, and obvious peritumoral edema, which suggests that

Table 3: Demographic data and conventional MR imaging characteristics of *BRAF p. V600E*-mutant and *BRAF p. V600E* wild-type APXA

	<i>BRAF p. V600E</i> - Mutant APXA	<i>BRAF p. V600E</i> Wild-Type APXA	<i>P</i> Value
Clinical data			
Male sex (No.) (%)	3 (60%)	3 (37.5%)	.413
Mean age (yr)	33 (SD, 13)	43 (SD, 6)	.284
Presenting symptoms (No.)			.850
Seizures	3	6	
Headache or increased ICP	1	1	
Neurologic deficit	1	1	
Location (No.)			.276
Frontal lobe	0	3	
Temporal lobe	2	3	
Occipital lobe	1	0	
Parietal lobe	2	1	
Insula	0	1	
Superficial location (No.) (%)	4 (80.0%)	7 (87.5%)	.641
Conventional MR imaging			
Mean size (cm)	3.2 (SD, 0.5)	3.3 (SD, 0.7)	.943
Presence of cystic degeneration (No.) (%)	3 (60%)	6 (75%)	.343
Peritumoral edema (mean) (cm)	1.8 (SD, 0.4)	2.4 (SD, 0.9)	.354
Heterogeneous enhancement (No.) (%)	5 (100%)	6 (75%)	.359
Leptomeningeal contact (No.) (%)	4 (80%)	7 (87.5%)	.641

Note:— ICP indicates intracranial pressure.

Tumor size (cm)



The presence and degree of peritumoral edema (cm)

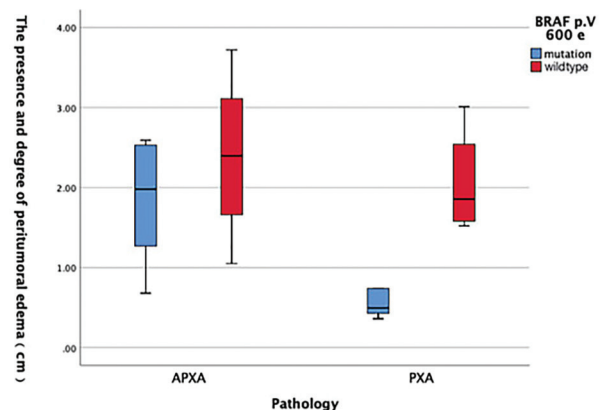


FIG 2. A significant difference ($P < .05$) between the *BRAF p. V600E*-mutant type PXA group and the *BRAF p. V600E* wild-type PXA group is demonstrated with both tumor size and the presence and degree of peritumoral edema. However, there was no significant difference in both tumor size and the maximum diameter of peritumoral edema between the *BRAF p. V600E*-mutant type APXA group and *BRAF p. V600E* wild-type APXA group.

BRAF p. V600E wild-type PXA may be more similar to high-grade gliomas in morphology. This conclusion may provide clinicians with strategies for tumor resection: For tumors identified by preoperative MR imaging such as *BRAF p. V600E* wild-type, more aggressive and expanded resection may be performed. For *BRAF p. V600E*-mutant tumors judged by preoperative MR imaging, a relatively conservative treatment strategy should be adopted, especially if the tumors are located in functional areas.

Table 4: Advanced MR imaging characteristics of the PXA and APXA groups

	PXA	APXA	P Value
ADCmin ($\times 10^{-6}$ mm ² /s)	951 (SD, 88)	755 (SD, 77)	<.001
ADCmean ($\times 10^{-6}$ mm ² /s)	1173 (SD, 159)	956 (SD, 106)	<.001

In the present study, we demonstrate that the rate of water diffusion of APXA, as reflected by the ADC ratio, was significantly lower than the rate of water diffusion of PXA, consistent with previous studies. Compared with *BRAF p. V600E*-mutant PXA, the ADC value of wild-type PXA was significantly lower. These results might be due to markedly high cellularity and a high nuclear/cytoplasmic ratio. Therefore, our findings reveal that ADC could be a useful imaging parameter for assessing the differences between *BRAF p. V600E*-mutant PXA and wild-type PXA. Only a few prior studies in the literature evaluated PXA using DWI.³⁷ The results of this study underline the usefulness of this simple imaging biomarker for radiogenomic correlation. However, future investigations are needed to better define the relationship between DWI features of *BRAF p. V600E*-mutant PXA and tumoral microarchitecture.

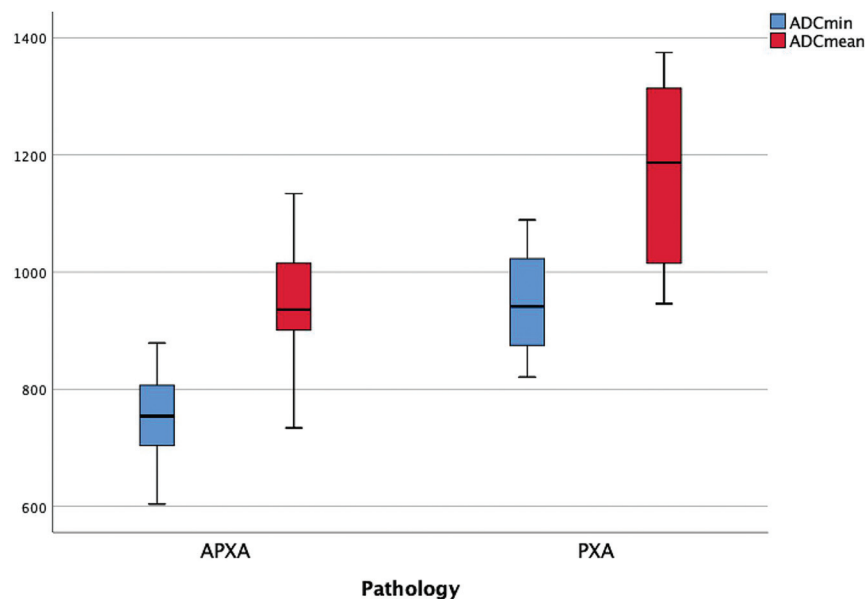


FIG 3. Boxplots representing ADCmin and ADCmean of PXA and APXA. A significant difference ($P < .05$) between PXA and APXA is demonstrated with both ADCmin and ADCmean.

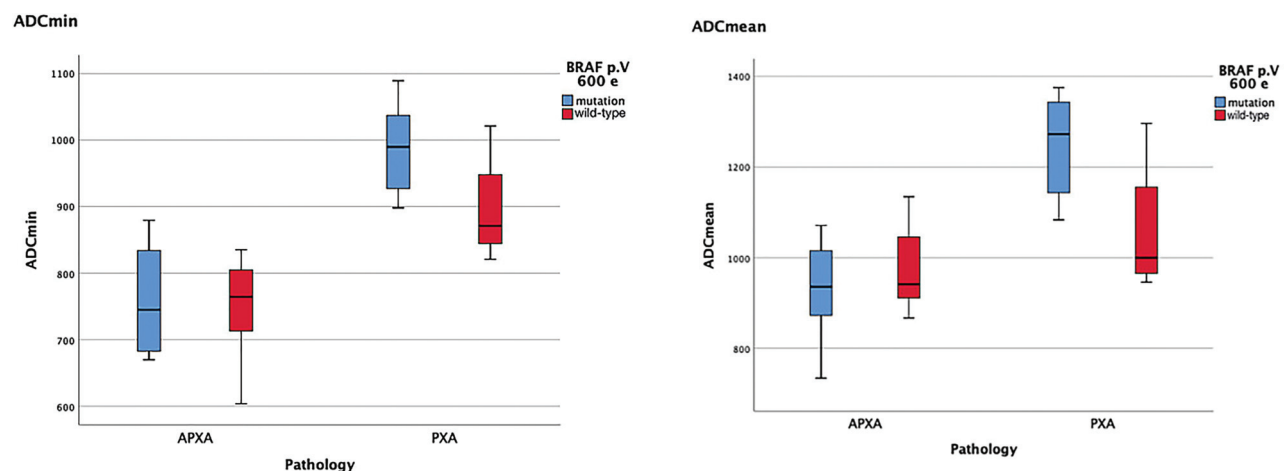


FIG 4. A significant difference ($P < .05$) between the *BRAF p. V600E*-mutant type PXA group and the *BRAF p. V600E* wild-type PXA group is demonstrated with both ADCmin and ADCmean. However, there was no significant difference in both ADCmin and ADCmean between the *BRAF p. V600E*-mutant type APXA group and *BRAF p. V600E* wild-type APXA group.

Table 5: Advanced MR imaging characteristics of the *BRAF p. V600E*-mutant PXA and *BRAF p. V600E* wild-type PXA groups

	<i>BRAF p. V600E</i> - Mutant PXA	<i>BRAF p. V600E</i> Wild- Type PXA	P Value
ADCmin ($\times 10^{-6}$ mm ² /s)	988 (SD, 73)	896 (SD, 86)	.047
ADCmean ($\times 10^{-6}$ mm ² /s)	1248 (SD, 116)	1060 (SD, 159)	.049

Table 6: Advanced MR imaging characteristics of the *BRAF p. V600E*-mutant APXA and *BRAF p. V600E* wild-type APXA groups

	<i>BRAF p. V600E</i> - Mutant APXA	<i>BRAF p. V600E</i> Wild- Type APXA	P Value
ADCmin ($\times 10^{-6}$ mm ² /s)	762 (SD, 92)	750 (SD, 73)	1
ADCmean ($\times 10^{-6}$ mm ² /s)	925 (SD, 131)	974 (SD, 92)	.724

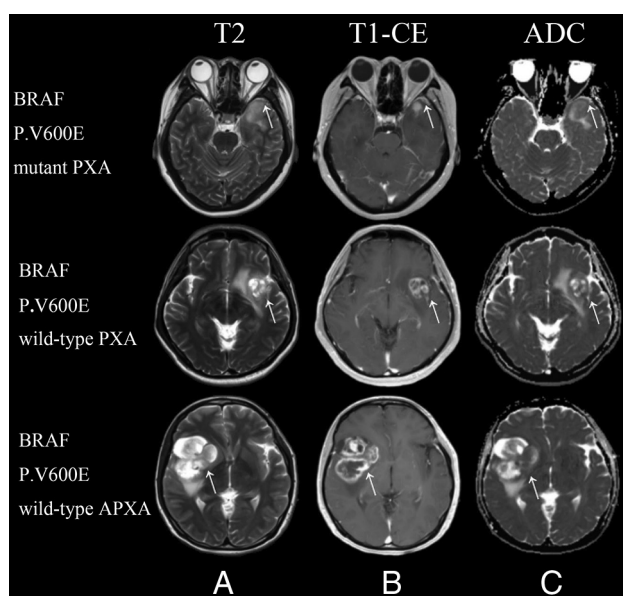


FIG 5. Conventional and advanced MR images of *BRAF p. V600E*-mutant and wild-type PXA. *Upper row:* A 27-year-old female patient with a *BRAF p. V600E*-mutant pleomorphic astrocytoma. T2WI (A) and contrast-enhanced T1WI (B) show a homogeneous contrast-enhancing solid tumor with mild perilesional edema located in the temporal lobe. C, A correlative ADC map shows the tumor with an elevated ADC value (ADCmin = 1089). *Second row:* A 53-year-old male patient with a *BRAF p. V600E* wild-type pleomorphic astrocytoma. T2WI (A) and contrast-enhanced T1WI (B) show a heterogeneous contrast-enhancing tumor with cystic degeneration and marked perilesional edema located in the insula. C, A correlative ADC map shows the lesion with a decreased ADC value (ADCmin = 875). *Third row:* A 46-year-old female patient with a *BRAF p. V600E* wild-type anaplastic pleomorphic astrocytoma. T2WI (A) and contrast-enhanced T1WI (B) show a heterogeneous contrast-enhancing tumor with cystic degeneration and marked perilesional edema located in the insula. C, A correlative ADC map shows the lesion with diffusion restriction (ADCmin = 704). CE indicates contrast-enhanced.

We did not find any MR imaging differences between the *BRAF p. V600E*-mutant APXA and the wild-type, whether using conventional or advanced imaging, possibly because the imaging features of APXA itself, similar to the imaging features of high-grade astrocytomas, mask the differences caused by *BRAF p.*

V600E-mutant. Vuong et al²⁰ previously proposed that *BRAF p. V600E* only showed a significant improvement in patient survival in low-grade tumors. These results illustrate the limitations of the *BRAF p. V600E*-mutant on MR imaging morphologic and clinical outcomes of APXA.

Several potential limitations of this study should be mentioned. First, our research is a single-center observational retrospective study. The influence caused by the deviation of the retrospective study cannot be excluded. Second, although the sample size included in this study is relatively large among the reports on PXA thus far, the number of patients in our study remained limited, whether it was PXA or APXA, *BRAF p. V600E*-mutant or wild-type. Therefore, the conclusions should be interpreted with caution. Third, because ADC values were not used to guide the biopsy in this retrospective study, it was not possible to evaluate the correlations between regions of lower ADC in PXA and hypercellularity point to point. These correlations should be performed in our future study. Larger multicenter prospective studies are necessary to perform an accurate analysis of the relationship between imaging features and genomic characteristics of this rare tumor.

CONCLUSIONS

BRAF p. V600E wild-type PXA can present with more aggressive conventional and advanced MR imaging features, mimicking high-grade astrocytomas at initial diagnosis, than *BRAF p. V600E*-mutant PXA. Greater maximum tumor diameter, heterogeneous contrast enhancement, obvious peritumoral edema, and lower ADCmin and ADCmean are more common features in *BRAF p. V600E* wild-type PXA compared with *BRAF p. V600E*-mutant PXA. Neurosurgeons should carefully interpret MR images before an operation in cases that may be considered PXA and select an appropriate surgical strategy according to the prediction of *BRAF p. V600E* genotype.

Disclosure forms provided by the authors are available with the full text and PDF of this article at www.ajnr.org.

REFERENCES

1. Rao AA, Laack NN, Giannini C, et al. **Pleomorphic xanthoastrocytoma in children and adolescents.** *Pediatr Blood Cancer* 2010;55:290–94 [CrossRef Medline](#)
2. Kepes JJ, Rubinstein LJ, Eng LF. **Pleomorphic xanthoastrocytoma: a distinctive meningocerebral glioma of young subjects with relatively favorable prognosis—a study of 12 cases.** *Cancer* 1979;44:1839–52
3. Kleihues P, Burger PC, Scheithauer BW. **The new WHO classification of brain tumors.** *Brain Pathol* 1993;3:255–68 [CrossRef Medline](#)
4. Louis DN, Ohgaki H, Wiestler OD, et al. **The 2007 WHO classification of tumors of the central nervous system.** *Acta Neuropathol* 2007;114:97–109 [CrossRef Medline](#)
5. Ida CM, Rodriguez FJ, Burger PC, et al. **Pleomorphic xanthoastrocytoma: natural history and long-term follow-up.** *Brain Pathol* 2015;25:575–86 [CrossRef Medline](#)
6. Hirose T, Ishizawa K, Sugiyama K, et al. **Pleomorphic xanthoastrocytoma: a comparative pathological study between conventional and anaplastic types.** *Histopathology* 2008;52:183–93 [CrossRef Medline](#)
7. Giannini C, Scheithauer BW, Burger PC, et al. **Pleomorphic xanthoastrocytoma: what do we really know about it?** *Cancer* 1999;85:2033–45

8. Louis DN, Perry A, Reifenberger G, et al. **The 2016 World Health Organization Classification of Tumors of the Central Nervous System: a summary.** *Acta Neuropathol* 2016;131:803–20 [CrossRef Medline](#)
9. Wesseling P, Capper D. **WHO 2016 classification of gliomas.** *Neuropathol Appl Neurobiol* 2018;44:139–50 [CrossRef Medline](#)
10. Marton E, Feletti A, Orvieto E, et al. **Malignant progression in pleomorphic xanthoastrocytoma: personal experience and review of the literature.** *J Neurol Sci* 2007;252:144–53 [CrossRef Medline](#)
11. Rutkowski MJ, Oh T, Niflioglu GG, et al. **Pleomorphic xanthoastrocytoma with anaplastic features: retrospective case series.** *World Neurosurg* 2016;95:368–74 [CrossRef Medline](#)
12. Ma C, Feng R, Chen H, et al. **BRAF V600E, TERT, and IDH2 mutations in pleomorphic xanthoastrocytoma: observations from a large case-series study.** *World Neurosurg* 2018;120:e1225–33 [CrossRef Medline](#)
13. Dias-Santagata D, Lam Q, Vernovsky K, et al. **BRAF V600E mutations are common in pleomorphic xanthoastrocytoma: diagnostic and therapeutic implications.** *PLoS One* 2011;6:e17948 [CrossRef Medline](#)
14. Tatevossian RG, Lawson AR, Forshew T, et al. **MAPK pathway activation and the origins of pediatric low-grade astrocytomas.** *J Cell Physiol* 2010;222:509–14 [CrossRef Medline](#)
15. Jacob K, Quang-Khuong DA, Jones DT, et al. **Genetic aberrations leading to MAPK pathway activation mediate oncogene-induced senescence in sporadic pilocytic astrocytomas.** *Clin Cancer Res* 2011;17:4650–60 [CrossRef Medline](#)
16. Maurer G, Tarkowski B, Baccarini M. **Raf kinases in cancer: roles and therapeutic opportunities.** *Oncogene* 2011;30:3477–88 [CrossRef Medline](#)
17. Schindler G, Capper D, Meyer J, et al. **Analysis of BRAF V600E mutation in 1,320 nervous system tumors reveals high mutation frequencies in pleomorphic xanthoastrocytoma, ganglioglioma and extra-cerebellar pilocytic astrocytoma.** *Acta Neuropathol* 2011;121:397–405 [CrossRef Medline](#)
18. Chamberlain MC. **Salvage therapy with BRAF inhibitors for recurrent pleomorphic xanthoastrocytoma: a retrospective case series.** *J Neurooncol* 2013;114:237–40 [CrossRef Medline](#)
19. Tabouret E, Bequet C, Denicolaï E, et al. **BRAF mutation and anaplasia may be predictive factors of progression-free survival in adult pleomorphic xanthoastrocytoma.** *Eur J Surg Oncol* 2015;41:1685–90 [CrossRef Medline](#)
20. Vuong HG, Altibi AM, Duong UN, et al. **BRAF mutation is associated with an improved survival in glioma—a systematic review and meta-analysis.** *Mol Neurobiol* 2018;55:3718–24 [CrossRef Medline](#)
21. Tonse R, Gupta T, Epari S, et al. **Impact of WHO 2016 update of brain tumor classification, molecular markers and clinical outcomes in pleomorphic xanthoastrocytoma.** *J Neurooncol* 2018;136:343–50 [CrossRef Medline](#)
22. Brown NF, Carter T, Kitchen N, et al. **Dabrafenib and trametinib in BRAFV600E mutated glioma.** *CNS Oncol* 2017;6:291–96 [CrossRef Medline](#)
23. She D, Liu J, Xing Z, et al. **MR imaging features of anaplastic pleomorphic xanthoastrocytoma mimicking high-grade astrocytoma.** *AJNR Am J Neuroradiol* 2018;39:1446–52 [CrossRef Medline](#)
24. Louis DN, Wesseling P, Aldape K, et al. **cIMPACT-NOW update 6: new entity and diagnostic principle recommendations of the cIMPACT-Utrecht meeting on future CNS tumor classification and grading.** *Brain Pathol* 2020;30:844–56 [CrossRef Medline](#)
25. Gimi B, Cederberg K, Derinkuyu B, et al. **Utility of apparent diffusion coefficient ratios in distinguishing common pediatric cerebellar tumors.** *Acad Radiol* 2012;19:794–800 [CrossRef Medline](#)
26. Mukherjee P, Miller JH, Shimony JS, et al. **Normal brain maturation during childhood: developmental trends characterized with diffusion-tensor MR imaging.** *Radiology* 2001;221:349–58 [CrossRef Medline](#)
27. Perkins SM, Mitra N, Fei W, et al. **Patterns of care and outcomes of patients with pleomorphic xanthoastrocytoma: a SEER analysis.** *J Neurooncol* 2012;110:99–104 [CrossRef Medline](#)
28. Yan J, Cheng J, Liu F, et al. **Pleomorphic xanthoastrocytomas of adults: MRI features, molecular markers, and clinical outcomes.** *Sci Rep* 2018;8:14275 [CrossRef Medline](#)
29. Yang RR, Aibaidula A, Wang WW, et al. **Pediatric low-grade gliomas can be molecularly stratified for risk.** *Acta Neuropathol* 2018;136:641–55 [CrossRef Medline](#)
30. Lindsay AJ, Rush SZ, Fenton LZ. **Pediatric posterior fossa ganglioglioma: unique MRI features and correlation with BRAF V600E mutation status.** *J Neurooncol* 2014;118:395–404 [CrossRef Medline](#)
31. Ostrom QT, Gittleman H, Liao P, et al. **CBTRUS Statistical Report: Primary brain and other central nervous system tumors diagnosed in the United States in 2010–2014.** *Neuro Oncol* 2017;19:v1–88 [CrossRef Medline](#)
32. Thakkar JP, Dolecek TA, Horbinski C, et al. **Epidemiologic and molecular prognostic review of glioblastoma.** *Cancer Epidemiol Biomarkers Prev* 2014;23:1985–96 [CrossRef Medline](#)
33. Rasmussen BK, Hansen S, Laursen RJ, et al. **Epidemiology of glioma: clinical characteristics, symptoms, and predictors of glioma patients grade I–IV in the Danish Neuro-Oncology Registry.** *J Neurooncol* 2017;135:571–79 [CrossRef Medline](#)
34. Yu S, He L, Zhuang X, et al. **Pleomorphic xanthoastrocytoma: MR imaging findings in 19 patients.** *Acta Radiol* 2011;52:223–28 [CrossRef Medline](#)
35. Crespo-Rodriguez AM, Smirniotopoulos JG, Rushing EJ. **MR and CT imaging of 24 pleomorphic xanthoastrocytomas (PXA) and a review of the literature.** *Neuroradiology* 2007;49:307–15 [CrossRef Medline](#)
36. Purkait S, Bansal S, Malgulkar PB. **BRAF V600E-mutated central nervous system tumor with divergent morphological feature: anaplastic pleomorphic xanthoastrocytoma-like and astroblastoma-like.** *Neuropathology* 2019;39:64–67 [CrossRef Medline](#)
37. Moore W, Mathis D, Gargan L, et al. **Pleomorphic xanthoastrocytoma of childhood: MR imaging and diffusion MR imaging features.** *AJNR Am J Neuroradiol* 2014;35:2192–96 [CrossRef Medline](#)